

European Journal of Pharmaceutics and Biopharmaceutics 50 (2000) 301-306

European

Journal of

Pharmaceutics and

Biopharmacentics

www.elsevier.com/locate/ejphabio

Research paper

Biopharmaceutical characterization of oral theophylline and aminophylline tablets. Quantitative correlation between dissolution and bioavailability studies

Jaleh Varshosaz*, Taghi Ghafghazi, Ahmad Raisi, Mahmood Falamarzian

Faculty of Pharmacy and Pharmaceutical Sciences, Isfahan University of Medical Sciences, Isfahan, Iran
Received 26 July 1999; accepted in revised form 25 January 2000

Abstract

Considering the narrow therapeutic index of theophylline and the low range between the safe and toxic serum concentrations of this drug, the study of its pharmacokinetic properties is necessary. However, considering the time consuming and expensive in vivo tests, quantitative correlation between in vivo bioavailability and in vitro dissolution tests can be used routinely in quality control tests of these drug products to predict the in vivo pharmacokinetic parameters. For this reason healthy human volunteers were used for in vivo studies and serum samples were analyzed by a fluorescence polarization immunoassay analysis (FPIA) method. The results showed that an open one compartmental model could best describe the pharmacokinetic properties of orally administered theophylline and aminophylline tablets. Linear regression analysis by least-square method showed a good correlation between some in vivo and in vitro parameters obtained from dissolution studies by rotating basket and paddle methods. $D_{30}\%$ (percentage of drug dissolved in vitro after 30 min) and $F_{0.5}\%$ (drug absorbed in vivo after half an hour calculated by Wagner–Nelson equation) showed best correlation (r = 0.99036). C_{max} (maximum serum concentration) of this drug also correlates well with $t_{25\%}$ (time required to dissolve 25% of the drug). The calculated correlation coefficients could best predict the actual values of some pharmacokinetic parameters; $AUC_{0\to\infty}$, $AUC_{0\to\infty}$. 2000 Elsevier Science B.V. All rights reserved.

Keywords: Bioavilability; Theophylline; Asthma; Pharmacokinetics; Dissolution

1. Introduction

Theophylline is a methylxanthine derivative with a narrow therapeutic index [1,2] and a serum concentration between $10-20~\mu g/ml$ is too effective and safe in controlling the signs of asthma and bronchospasms caused by other diseases.

As the induced bronchodilation is diminished by reducing its serum concentration and on the other hand its toxicity risk is increased above 20 μ g/ml, the fluctuations of its serum concentrations produce some clinically important problems. In other words, there is a very close relationship between plasma concentration of this drug, its toxic and therapeutic effects [3,4]. Therefore, it seems that plasma drug monitoring is quite necessary for different dosage forms of theophylline.

Since drug release pattern from each dosage form is significantly affected by the special design used in manu-

facturing the formulations, planning an in vitro dissolution method corresponding to the in vivo drug absorption rate, will facilitate the development of the drug formulations and quality control tests. From the economic, technical, and ethical perspectives of bioavailability studies, it is necessary to establish an in vitro test method that can predict the progress of drug release and the absorption of products in vivo [5].

El-Yazigi and Sawchuk [6] found good correlations between bioavailability parameters and dissolution characteristics of theophylline sustained-release dosage forms in rabbit. Ritschel et al. [7] also reported such correlations in beagle dog. Brockmeir [8], Hwan Chung and Shim [9] also compared in vitro dissolution rates with in vivo parameters in human using theophylline saliva concentrations.

The objective of this study was to correlate logically the rapid simple and practical in vitro dissolution methods of theophylline tablets with its in vivo human pharmacokinetic parameters. This correlation will be able to predict the bioavailability of theophylline dosage forms instead of difficult, time-consuming and expensive in vivo tests routinely.

^{*} Corresponding author. School of Pharmacy and Pharmaceutical Services, Mezar Jerib Ave., P.O. Box 81745-359, Isfahan, Iran. Tel.: +98-31-688-064; fax:98-31-688-065.

2. Materials and methods

2.1. Materials

All studied dosage forms were purchased commercially; 100 mg enteric coated tablet of aminophylline (ethylenediamine salt of theophylline) (Iran Hormone, Iran) with code A, 100 mg sugar coated tablets of aminophylline (Abidi laboratories, Iran) with code B, 100 mg tablets of aminophylline (Artezan, Germany) with code C, and 200 mg tablets of theophylline (Abidi Laboratories, Iran) with code D, anhydrous theophylline and aminophylline powder (Bohringer, Germany).

2.2. Methods

2.2.1. Bioavailability studies

Six healthy male volunteers, ranging 21–24 years old and weighing 58–75 kg, participated in this study. The subjects were not smokers and had not taken any drugs during the preceding 2 months. They were found to be normal on physical examination, a blood screen and urinalysis. None of them had a history of any serious or chronic disease. They were allowed no xanthine-containing beverages for 3 days before each administered dose and for the duration of the sampling schedule. No food was permitted for 3 h after taking the tablets. Each volunteer received a 200 mg single dose of products A, B, C and D in a complete cross-over design. The interval between each studied dosage form was 2 weeks. The respective dose of the drug was given with 200 ml of water.

2.2.2. Sample collection

Three milliliters of blood was taken from the forearm vein just prior to dosing and at 0.5, 1, 2, 3, 5, 7, and 9 h after dosing. Each blood sample was allowed to stand for about 1 h and then serum was separated by centrifugation. The serum samples were kept frozen (-20° C) until the time of analysis.

2.2.3. Assay

The serum theophylline concentrations were determined by a TD_x analysis (TD_x^{\circledast} System Analyzer AbbottTM Laboratories, Dallas, TX) based on a fluorescence polarization immunoassay analysis (FPIA) method [10].

2.2.4. Bioavailability parameters

The area under the serum concentration-time curve from time zero to time t (AUC_{0→t}) was calculated using the linear trapezoidal rule. The area from time t to infinity was estimated by C_t/K_{el} , where C_t is the serum theophylline concentration observed at time t, and K_{el} is the apparent elimination rate constant of theophylline obtained from the slope of log-linear portion of the curve by least square regression analysis. Moreover, the area from time zero to infinity was calculated by:

$$AUC_{0\to\infty} = AUC_{0\to t} + C_t/K_{el} \tag{1}$$

The fraction of the dose absorbed in time t (F_t) was calculated from the Wagner-Nelson equation [11].

$$F_t = \frac{(C_t + K_{el} \cdot AUC_{0 \to \infty})}{K_{el} \cdot AUC_{0 \to \infty}}$$
 (2)

2.2.5. Statistical analysis

A one-way ANOVA post hoc multiple comparison LSD test with P < 0.05 was used for statistical analysis using SPSS computer software. The correlation coefficients of the regression lines between in vitro dissolution parameters and in vivo parameters were obtained by the linear least square method. The correlation coefficients were examined for their significance with the t-test.

3. Results

3.1. Bioavailability results

The results of bioavailability and pharmacokinetic studies of four theophylline and aminophylline tablets; A, B, C and D in six volunteers are shown in Table 1. Fig. 1 indicates that an open one compartmental model best describes the disposition of oral theophylline and its ethylenediamine derivative in human body [9,12–14]. A great difference can be seen between the absorption (K_a) and elimination rate $(K_{\rm el})$ of this drug (Table 1). The average elimination half life $(T_{1/2})$ in six volunteers showed no significant difference between different studied products (Table 1). The differences between bioavailability (expressed as the area under the time-concentration curve until infinity, i.e. $AUC_{0\to\infty}$) of products A and C (P < 0.01) and also between C and D (P < 0.05) were significant so that C was greater than A and D (Table 1). The results of in vitro dissolution tests also showed that the dissolution rate of C is faster than others [15].

4. Discussion

The great difference between K_a and K_{el} of the studied products shows that theophylline absorbs rapidly from the studied dosage forms while its elimination happens slowly. However, as it shows a one-compartmental pharmacokinetic model (Fig. 1), it may be concluded that its elimination rate from the central compartment (i.e. blood) is slow [9,12]. This suggests the necessity of constant monitoring of this drug in the blood of the patients under long treatment with theophylline to prevent any intoxication.

However, the great variations between $T_{1/2}$ in different subjects probably indicates the interindividual differences in hepatic metabolism rate of theophylline [14,16–19]. Different types of sustained release theophylline dosage forms also show a great interindividual variation in extent and absorption rate [20].

Pharmacokinetic data (mean ± SD) of four commercial products of theophylline^a

Product	Product $C_{\text{max}}^{\text{b}}(\mu \text{g/ml})$ $T_{\text{max}}^{\text{c}}(\text{h})$	$T_{\mathrm{max}}^{}\mathrm{c}}(\mathrm{h})$	$T_{1/2}^{d}(h)$	$K_{\rm a}^{\rm e} \left({\rm h}^{-1} \right)$	$K_{\rm a}^{\rm f} \left({\rm h}^{-1} \right)$	$AUC_{0\rightarrow 0.5}^g \; (\mu g/ml \cdot h)$	$AUC_{0\rightarrow 1}^{ h} \; (\mu g/ml \cdot h)$	$AUC_{0\rightarrow1}{}^{h}\left(\mu g/ml\cdot h\right) AUC_{0\rightarrow9}{}^{i}\left(\mu g/ml\cdot h\right) AUC_{0\rightarrow\infty}{}^{j}\left(\mu g/ml\cdot h\right) F_{0.5}\%^{k}$	$\mathrm{AUC}_{0\to\infty}^{j}(\mu g/m l\cdot h)$	$F_{0.5}\%^{\rm k}$
A	6.364 ± 0.587	2.213 ± 0.447	7.985 ± 0.769	1.532 ± 0.853	0.087 ± 0.008	0.771 ± 0.339	2.714 ± 1.040	40.944 ± 3.786	80.687 ± 4.945	44.48 ± 17.4
В	7.045 ± 0.771	2.000 ± 0.707	8.176 ± 1.680	1.484 ± 0.819	0.088 ± 0.018	0.425 ± 0.495	2.110 ± 1.420	44.639 ± 3.576	90.707 ± 18.114	21.96 ± 24.4
C	6.886 ± 0.970	1.900 ± 0.894	8.448 ± 1.420	1.689 ± 0.811	0.084 ± 0.012	0.939 ± 0.719	3.397 ± 1.654	45.227 ± 2.774	93.982 ± 6.965	47.46 ± 31.3
О	6.040 ± 0.509	2.000 ± 0.707	7.308 ± 2.683	1.192 ± 0.459	0.105 ± 0.037	0.562 ± 0.336	2.354 ± 1.014	36.512 ± 6.443	70.907 ± 26.595	32.90 ± 16.7

^a Aminophylline tablets (A–C) and theophylline tablet (D) (n=6).

^b Maximum serum concentration.
^c Time required to reach maximum serum concentration.
^d Elimination half life.

^e Absorption rate constant. ^f Elimination rate constant.

 $^{\rm g}$ Area under the serum concentration-time curve from zero time to 0.5 h. Area under the serum concentration-time curve from zero time to 1 h.

 $^{\rm i}$ Area under the serum concentration-time curve from zero time to 9 h. $^{\rm j}$ Area under the serum concentration-time curve from zero time to infinity. $^{\rm k}$ Percentage of drug absorbed after half an hour.

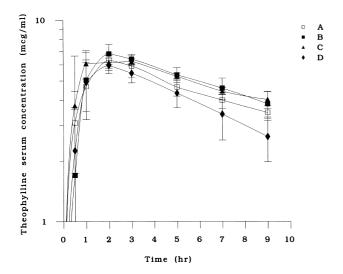
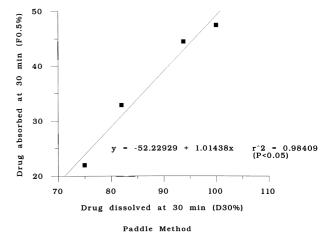


Fig. 1. Mean log of theophylline serum concentrations of human healthy volunteers after oral administration of a single dose (equal to 200 mg of pure theophylline) from aminophylline (A–C) and theophylline (D) tablets (n = 6).

Similar results for $T_{\rm max}$ of the studied products suggests that there may be a problem in the formulation of product A. As this tablet is an enteric coated form, it needs a 3.61 ± 1.47 h to be evacuated from the stomach [13]. In vitro dissolution studies also showed the rapid release of the drug from tablet A [15].

4.1. Quantitative correlation of bioavailability and dissolution parameters

Some bioavailability parameters showed a good degree of correlation with some dissolution parameters. For example, Fig. 2 shows a very high correlation ($r=0.990,\,P<0.05$ for paddle method and $r=0.984,\,P<0.05$ for rotating basket method) between the fraction absorbed in 0.5 h calculated by Wagner–Nelson Eq. ($F_{0.5}\%$) and the percentage dissolved in 30 min ($D_{30}\%$) for four products. Some



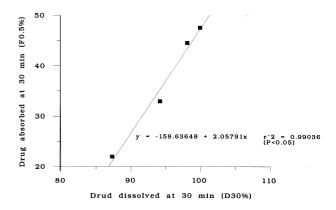


Fig. 2. Quantitative correlation between percentage of the ophylline absorbed ($F_{0.5}\%$) and dissolved ($D_{30}\%$) after 30 min by two methods of dissolution studies (Rotating Basket and Paddle Method).

pairs of parameters showed significant correlations, as shown in Table 2. The percentage of dissolved drug from studied dosage forms by rotating basket method showed better correlations with some serum bioavailability parameters than did paddle method, except in one case, i.e. the correlation coefficient between $F_{0.5}\%$ and $D_{30}\%$ which was greater by the paddle method (Table 2). Using the equations

Table 2 Correlation coefficients between in vitro dissolution and in vivo pharmacokinetics parameters

Dissolution test method	In vitro parameter	In vivo parameter	Correlation coefficient (r)	Slope	y-Intercept
I^a	$D_{30}\%$	F _{0.5} %	0.984	1.014	-52.229
Π_p	$D_{30}\%$	$F_{0.5}\%$	0.990	2.058	-158.636
I	$D_{30}\%$	$AUC_{0\rightarrow 1}$	0.956	0.047	-1.504
П	$D_{30}\%$	$AUC_{0\rightarrow 1}$	0.882	0.880	-5.707
I	$D_{60}\%$	$AUC_{0\rightarrow 1}$	0.797	0.079	-4.853
П	$D_{60}\%$	$AUC_{0\rightarrow 1}$	0.669	0.137	-10.788
I	$D_{60}\%$	$AUC_{0\rightarrow\infty}$	0.693	1.287	-37.492
П	$D_{60}\%$	$AUC_{0\rightarrow\infty}$	0.680	2.598	-170.327
I	$t_{25\%}$	$C_{ m max}$	0.699 ^c	0.053	6.255
II	t _{25%}	$C_{ m max}$	0.701°	0.061	6.254

a Rotating Basket method.

b Paddle method.

^c Not significantly different.

Relationship between in vivo and in vitro parameters (I, Rotating Basket and II, Paddle method) of theophylline products to predict some pharmacokinetic parameters

Product	$D_{30}\%^{ m a}$		Actual $F_{0.5}\%^{\mathrm{b}}$	Predicted $F_{0.5}\%$		Actual AUC _{0→1} ^c (μg/ml·h)	Predicted (μg/ml·h)	redicted AUC _{0→1} µg/ml·h)	$D_{60}\%^{ m d}$		Actual $AUC_{0\rightarrow\infty}^{e}$ (µg/ml·h)	Predictec (µg/ml·h)	Predicted AUC _{0→∞} (μg/ml·h)	t _{25%} f (min)	Actual C _{max} ^g I (μg/ml)	Predicted C _{max} (µg/ml)
	I	П		I	П		I	II	I	П		I	П	Ι		
Ah	93.78	98.14	44.48		43.33	2.71	2.93	2.93		100	80.69	88.41	89.53	4.8	6.36	6.51
\mathbf{B}_{p}	74.94	87.32	21.96		21.05	2.11	2.04	1.97		97.37	90.71	81.49	82.69	15	7.04	7.06
ڻ ت	100	100	47.46	49.21	47.16	3.40	3.23	3.09	100	100	93.98	91.26	89.53	3.1	68.9	6.42
Ωp	81.96	94.22	32.9		35.26	2.35	2.37	2.58		94.26	70.91	75.13	74.61	1.6	6.04	6.34

^a Percentage of drug dissolved after 30 min.

^b Percentage of drug absorbed after half an hour.
^c Area under the serum concentration-time curve from zero time to 1 h.

^d Percentage of drug dissolved after 60 min.

 $^{\circ}$ Area under the serum concentration-time curve from zero time to infinity. Time required to dissolve 25% of the drug.

 $^{\rm g}$ Maximum serum concentration. $^{\rm h}$ Aminophylline tablets (A–C) and theophylline tablet (D).

obtained from the linear regression quantitative correlation between in vitro and in vivo parameters denoted in Table 2, the amount of in vitro parameters obtained by each dissolution study method were fitted in each equation of Table 2 and the amount of in vivo parameters were predicted (Table 3). For example, the mean amounts of $D_{30}\%$ for product A by rotating basket method and paddle method were 93.778 and 98.142%, respectively, which were fitted in the equations obtained from the correlation studies between $F_{0.5}\%$ and $D_{30}\%$ (Table 2 and Fig. 2), i.e. y = 1.014 x - 52.229(rotating basket method) and y = 2.058 x - 158.636 (paddle method), and the amount of $F_{0.5}\%$ was predicted by each of the two methods. This gives 42.90% for rotating basket method and 43.33% for paddle method (Table 3). As the studied products are immediate release and not sustainedrelease products, it is not surprising that about 44% of the administered dose is absorbed during the first 30 min of the administration. Chung and Shim [9] also reported a $T_{\rm max}$ of 1.30 \pm 0.98 h for a commercial rapid release theophylline dosage form in which 87% of the drug was absorbed after 1 h.

In this way as shown in Table 3, $D_{30}\%$ was used for prediction of $F_{0.5}\%$ and $AUC_{0\rightarrow 1}$, $D_{60}\%$ for prediction of $AUC_{0\rightarrow \infty}$, and $t_{25\%}$ for prediction of C_{max} . The amount of $D_{30}\%$, $D_{60}\%$, and $t_{25\%}$ for each product are shown in Table 3. The predicted amounts were too close to the actual values (Table 3). Therefore, it seems that the two methods of in vitro dissolution tests of theophylline solid dosage forms i.e., rotating basket and paddle method [15] are capable in predicting the in vivo pharmacokinetic parameters in human.

5. Conclusions

In conclusion, it was found that some blood bioavailability parameters of theophylline dosage forms, like $F_{0.5}\%$, $AUC_{0\rightarrow 1}$, $AUC_{0\rightarrow \infty}$ and C_{\max} may be predicted from in vitro dissolution data, like $D_{30}\%$, $D_{60}\%$ and $t_{25\%}$, by considering the correlation among in vitro and in vivo studies. It seems convenient and advantageous to use $D_{30}\%$ or $D_{60}\%$ instead of blood data, which needs frequent venepuncture for sampling, in the development, evaluation and quality control of theophylline dosage forms.

Both methods of dissolution studies of four commercial tablets of theophylline and aminophylline, i.e. rotating basket and paddle method were capable to predict the pharmacokinetic parameters of this drug in human blood which may be routinely used in quality control laboratories to omit the time consuming and difficult in vivo bioavailability studies in human. This correlation was best shown between the drug absorbed after 30 min $(F_{0.5}\%)$ and the drug dissolved after the same time.

References

- H.J. Myron, M. Robertand, K. Gerald, Clinical experience with theophylline. Relationships between dosage, serum concentration and toxicity, J. Am. Med. Assoc. 235 (18) (1976) 1983–1986.
- [2] R.E. Notari, Biopharmaceutics and clinical pharmacokinetics, an introduction, 4th ed., Marcel Dekker, Inc, New York, 1987 183/ 388/180.
- [3] E.G. Ellis, R. Koysooko, G. Levy, Pharmacokinetics of theophylline in children with asthma, Pediatrics 58 (1976) 542.
- [4] G.M.R. Vandenbossche, R.A. Lefebvre, A.G. DE Wilde, J.P. Remon, Performance of a modified starch hydrophillic matrix for the sustained release of theophylline in healthy volunteers, J. Pharm. Sci. 81 (3) (1992) 245–248.
- [5] L. Shan-Yang, K. Yuh-Horng, C. Hsiao-Ning, Preliminary evaluation of the correlation between in vitro release and in vivo bioavailability of two aminophylline slow-release tablets, J. Pharm. Sci. 79 (4) (1990) 326–330.
- [6] A. El-Yazigi, R.J. Sawchuk, In vitro-in vivo correlation and dissolution studies with oral theophylline dosage forms, J. Pharm. Sci. 74 (2) (1985) 161–164.
- [7] W.A. Ritschel, H.P. Koch, G.J. Alcorn, In vitro-in vivo correlation with SR theophylline preparations, Methods Find. Exp. Clin. Pharmacol. 6 (10) (1984) 609–618.
- [8] D. Brockmeir, In vitro-in vivo correlation, a time scaling problem, Evaluation of mean times, Arzneim.- Forsch 34 (1984) 1604–1607.
- [9] B.H. Chung, C.K. Shim, Dissolution of the ophylline from sustainedrelease dosage forms and correlation with saliva bioavailability parameters, J. Pharm. Sci. 76 (10) (1987) 784–787.
- [10] M.E. Jolley, S.D. Stroupe, K.S. Schwenzer, C.J. Wang, M.Lu. Steffes, H.D. Hill, D.M. Kelso, Fluorescence polarization immunoassay, an automated system for therapeutic drug determination, Clin. Chem. 27 (9) (1981) 1575–1579.
- [11] J.G. Wagner. Fundamentals of Clinical Pharmacokinetics, 1st ed., Drug Intelligence, Hamilton, IL, 1975, pp. 174–176.
- [12] L. Shan-Yang, Y. Juei-Chyi, Bioavailability studies of theophylline ethylcellulose microcapsules prepared by using ethyl-vinyl acetate copolymer as a coacervation inducing agent, J. Pharm. Sci. 76 (3) (1987) 219–223.
- [13] K.J. Simons, E.M. Frith, E.R. Simons, Dissolution and bioavailability studies of whole and halved sustained-release theophylline tablets, J. Pharm. Sci. 71 (5) (1982) 505–511.
- [14] R.A. Upton, J.F. Thiercelin, S. Riegelman, Intraindividual variability in theophylline pharmacokinetics: statistical verification in 39 of 60 healthy young adults, J. Pharmacok. Biopharm. 10 (1) (1982) 123– 133
- [15] J. Varshosaz, T. Ghafghazi, A. Raisi, M. Falamarzian, Study the dissolution rate of some theophylline and aminophylline tablets by rotating basket and paddle method, Res. Bull. Isfahan Univ. (2000) in press.
- [16] E.G. Ellis, R. Koysooko, G. Levy, Pharmacokinetics of theophylline in children with asthma, Pediatrics 58 (1976) 542.
- [17] P.A. Mitenko, R.I. Ogilvie, Rational intravenous doses of theophylline, N. Engl. J. Med. 289 (1973) 600–603.
- [18] P. Rangsithienchai, R.W. Newcomb, Aminophylline therapy in children: guidelines for dosage forms, J. Pediatr. 91 (2) (1977) 325–330.
- [19] J.G. Wagner, Intrasubject variation in elimination half lives of drugs which are appreciably metabolized, J. Pharmaco. Biopharm. 1 (1973) 165–173.
- [20] J.G. Hardman, L.E. Limbird, P.B. Molinoff, R.W. Ruddon, A. Goodman, The Pharmacological Basis of Therapeutics, Mc Graw-Hill, New York, 1996 p. 676.